



## VALIDATION DATA

# 1. Introduction

Olink® Organ Damage is a reagent kit measuring 92 immune response related human protein biomarkers simultaneously. The analytical performance of the product has been carefully validated and the results are presented below.

### 1.1 TECHNOLOGY

The Olink reagents are based on the Proximity Extension Assay (PEA) technology<sup>1,2</sup>, where 92 oligonucleotide labeled antibody probe pairs are allowed to bind to their respective target protein present in the sample. A PCR reporter sequence is formed by a proximity dependent DNA polymerization event, amplified, and subsequently detected and quantified using real-time PCR. The assay is performed in a homogeneous 96-well format without any need for washing steps, see Figure 1.

### 1.2 QUALITY CONTROLS

Internal and external controls have been developed by Olink for data normalization and quality control purposes. These controls have been designed to enable monitoring of the technical assay performance, as well as the quality of individual samples, providing information at each step of the Olink protocol (see Figure 1). The internal controls are added to each sample and include two Immunoassay controls, one Extension control and one Detection control. The Immunoassay controls (two non-human proteins) monitor all three steps starting with the immunoreaction. The Extension Control (an antibody linked to two matched oligonucleotides for immediate proximity independent of antigen binding) monitors the extension and readout steps and is used for data

normalization across samples. Finally, the Detection control (a synthetic double-stranded template) monitors the readout step. Samples for which one or more of the internal control values deviate from a pre-determined range will be flagged and may be removed before statistical analysis.

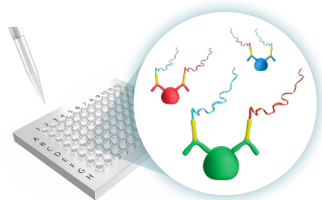
An external control, inter-plate control (IPC), is included on each plate and used in a second normalization step. This control is made up of a pool of probes similar to the Extension control (Ext Ctrl), but generated with 92 matching oligonucleotide pairs. Furthermore, the improves inter-assay precision and allows for optimal comparison of data derived from multiple runs. The term “Normalized Protein eXpression (NPX)” refers to normalized data as described above.

### 1.3 DATA ANALYSIS

Data analysis was performed by employing a pre-processing normalization procedure. For each sample and data point, the corresponding Cq-value for the Extension control was subtracted, thus normalizing for technical variation within one run. Normalization between runs is then performed for each assay by subtracting the corresponding dCq-value for the Interplate Control (IPC) from the dCq-values generated. In the final step of the pre-processing procedure the values are set relative to a correction factor determined by Olink. The generated Normalized Protein eXpression (NPX) unit is on a log2 scale where a larger number represents a higher protein level in the sample, typically with the background level at around zero. Linearization of data is performed by the mathematical operation  $2^{NPX}$ . Coefficient of variation (CV) calculations were performed on linearized values.

#### IMMUNOASSAY

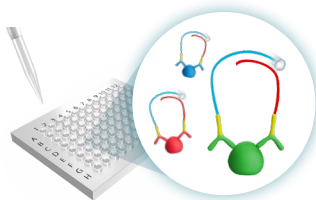
Allow the 92 antibody probe pairs to bind to their respective proteins in your samples.



Immunoassay control

#### EXTENSION

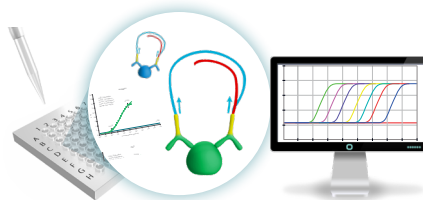
Extend and pre-amplify 92 unique DNA reporter sequences by proximity extension.



Extension control

#### DETECTION

Quantify each biomarker's DNA reporter using high throughput real-time qPCR.



Detection control

**Fig 1.** Olink assay procedure (above) and controls (below). The internal controls enables monitoring of the three core steps in the Olink assay and used for quality control and data normalization. Read out is performed by using the Fluidigm® Biomark™ or the Fluidigm® Biomark™ HD system.

## 2. Performance characteristics

### 2.1 SAMPLE TYPES

The ability to use different sample types was evaluated with Olink Organ Damage by collecting matched serum, EDTA, acid citrate dextrose (ACD), and sodium heparin plasma samples from 4 healthy individuals. Table 1 summarizes response values for 22 normal EDTA plasma samples expressed in NPX, as well as relative differences as compared to EDTA plasma. Variations observed between responses in heparin, citrate plasma and serum, as compared to EDTA plasma, were generally small, and all assays will therefore function without limitation in these sample types.

### 2.2 ANALYTICAL MEASUREMENT

#### DETECTION LIMIT

Calibrator curves were determined for 90 out of 92 biomarkers simultaneously in a multiplex format. Two protein biomarkers (RASA1 and ATP6AP2) lacked accessible recombinant antigen. Limit of detection (LOD) was defined as 3 standard deviations above background and reported in pg/mL for all assays where recombinant protein antigen was available, see Table 1 and Figure 2.

#### HIGH DOSE HOOK EFFECT

The high dose hook effect is a state of antigen excess relative to the reagent antibodies, resulting in falsely lower values. In such cases, a significantly lower value can be reported which leads to misinterpretation of results. Therefore, the hook effect was determined for each analyte, here reported in pg/mL for 91 out of 92 assays, see Table 1.

#### MEASURING RANGE

The analytical measuring range was defined by the lower limit of quantification (LLOQ) and upper limit of quantification (ULOQ) and reported in order of log<sub>10</sub>, see Table 1. The upper and lower limits of quantification, ULOQ and LLOQ, respectively were calculated with the following trueness and precision criteria; relative error  $\leq 30\%$  and CV  $\leq 30\%$ , of back-calculated values, and reported in pg/mL, see Table 1.

Three assays with their analytical data are shown in Figure 2 and the distribution of measuring ranges of 90 assays and endogenous plasma levels are shown in Figure 3. Separate calibrator curves established for each assay may be viewed at [www.olink.com](http://www.olink.com).

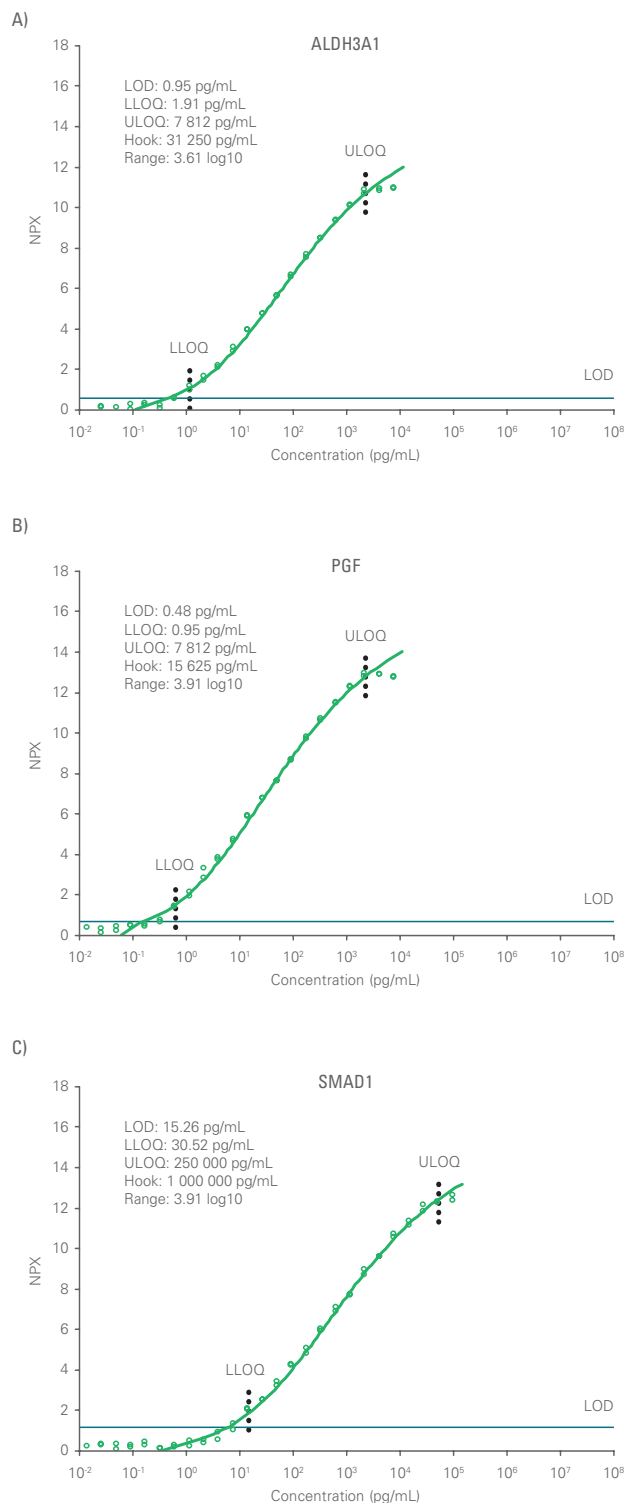
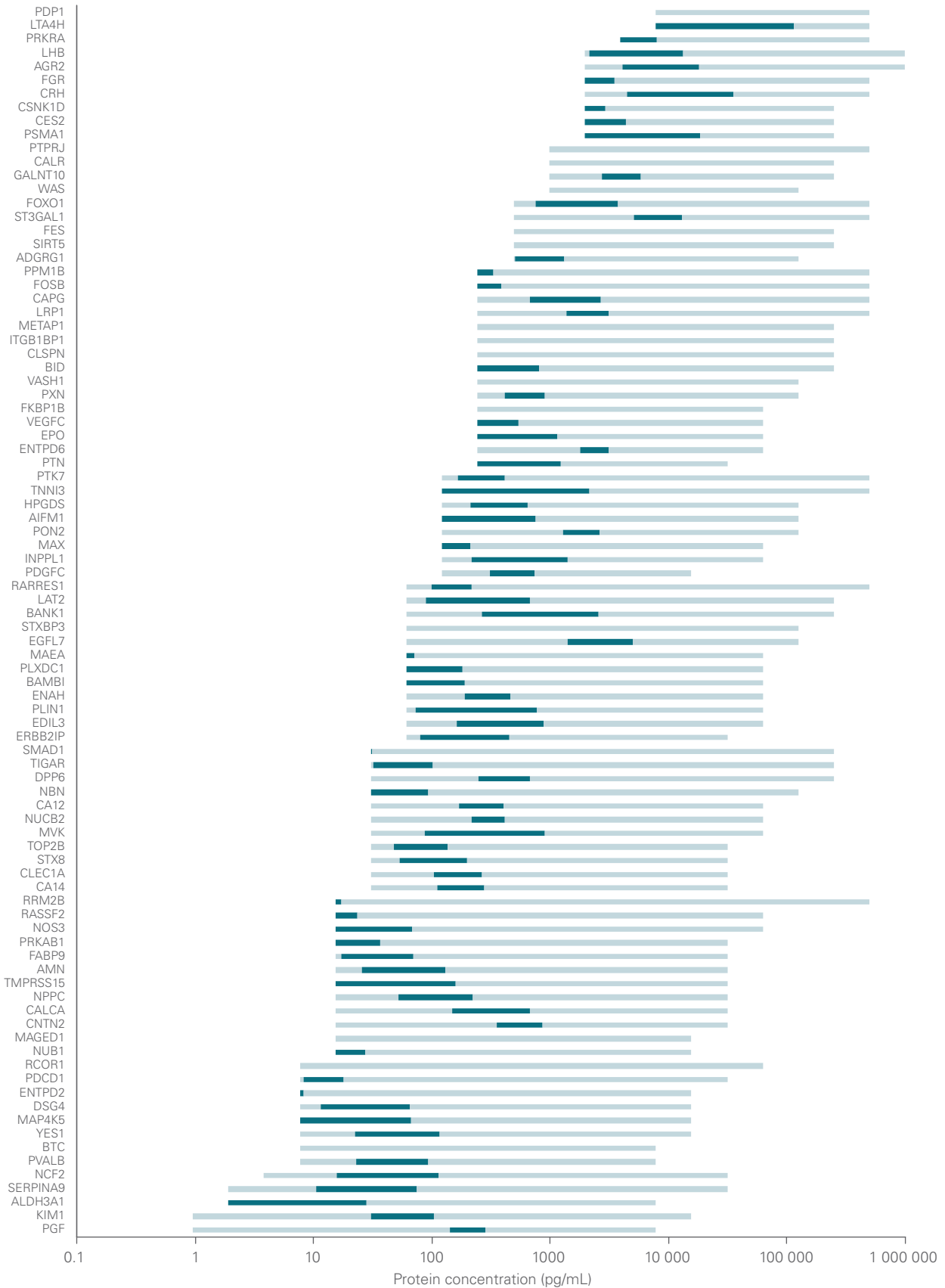


Fig 2. Calibrator curves from 3 assays and their corresponding analytical measurement data.



**Fig 3.** Distribution of analytical measuring range, defined by the lower and upper limits of quantification (LLOQ-ULOQ). Normal plasma levels (dark green bars) are denoted for 90 out of 92 analytes.

**Table 1.** Sample Types; Normalized Protein eXpression (NPX), Endogenous Interference, Analytical Measurement; Limit of Detection (LOD), Lower Limit of Quantification (LLOQ), Upper Limit of Quantification (ULOQ), High Dose Effect (Hook), Range and Precision indicative of assay performance are shown for 92 analytes. Not available, NA

Target	UniProt No	Sample types						Endogenous Interference	Analytical measurement				Precision		
		Normal plasma levels (NPX)			Relative to EDTA plasma (%)			(mg/mL)	pg/mL			log10	% CV		
		10th %tile	Median	90th %tile	ACD	Heparin	Serum	Hemolysate	LOD	LLOQ	ULOQ	Hook	Range	Intra	Inter
5'-AMP-activated protein kinase subunit beta-1 (PRKAB1)	Q9Y478	0.4	0.8	1.7	103	100	102	15	15	15	31 250	1 000 000	3.3	5	16
Adhesion G-protein coupled receptor G1 (ADGRG1)	Q9Y653	1.3	1.7	2.5	132	99	115	15	244	488	124 999	250 000	2.4	5	25
Aldehyde dehydrogenase, dimeric NADP-preferring (ALDH3A1)	P30838	0.5	1.9	3.6	111	95	111	15	0.95	1.91	7812	31 250	3.6	4	18
Anterior gradient protein 2 homolog (AGR2)	Q95994	1.6	2.3	3.8	124	101	117	15	976	1953	1 000 000	1 000 000	2.7	6	26
Apoptosis-inducing factor 1, mitochondrial (AIFM1)	Q95831	NA	1.2	2.7	99	89	90	15	122	122	124 999	1 000 000	3.0	6	21
B-cell scaffold protein with ankyrin repeats (BANK1)	Q8NDB2	2.5	4.2	6.0	34	108	34	15	61	61	250 000	499 999	3.6	6	22
BH3-interacting domain death agonist (BID)	P55957	NA	1.0	1.6	NA	NA	NA	0.5	244	244	250 000	1 000 000	3.0	5	26
BMP and activin membrane-bound inhibitor homolog (BAMBI)	Q13145	0.8	1.4	1.8	132	101	128	15	31	61	62 499	124 999	3.0	5	25
Calcitonin (CALCA)	P01258	4.2	5.4	6.8	116	113	116	15	7.63	15	31 250	124 999	3.3	5	18
Calreticulin (CALR)	P27797	0.4	1.0	1.3	NA	NA	NA	0.5	488	976	250 000	1 000 000	2.4	7	25
Carbonic anhydrase 12 (CA12)	Q43570	2.2	2.7	3.3	127	105	121	15	31	31	62 499	124 999	3.3	5	21
Carbonic anhydrase 14 (CA14)	Q9ULX7	1.8	2.2	2.7	114	101	115	15	31	31	31 250	31 250	3.0	4	16
Casein kinase I isoform delta (CSNK1D)	P48730	NA	0.6	1.0	NA	NA	NA	15	1953	1953	250 000	499 999	2.1	4	19
Claspin (CLSPN)	Q9HAW4	NA	NA	0.6	NA	NA	NA	15	122	244	250 000	499 999	3.0	8	16
CMP-N-acetylneuraminate-beta-galactosamide-alpha-2,3-sialyltransferase 1 (ST3GAL1)	Q11201	3.7	4.2	5.0	143	104	133	15	488	488	499 999	499 999	3.0	4	23
Cocaine esterase (CES2)	Q00748	0.5	1.2	1.8	107	93	96	15	1953	1953	250 000	1 000 000	2.1	5	18
Contactin-2 (CNTN2)	Q02246	4.0	4.5	5.1	138	105	133	15	15	15	31 250	31 250	3.3	4	19
Corticoliberin (CRH)	P06850	2.1	3.1	5.3	128	98	120	15	1953	1953	499 999	499 999	2.4	5	24
C-type lectin domain family 1 member A (CLEC1A)	Q8NC01	2.6	3.2	3.9	152	107	144	15	31	31	31 250	124 999	3.0	5	21
C-type natriuretic peptide (NPPC)	P23582	2.6	3.8	5.7	98	116	107	15	15	15	31 250	31 250	3.3	6	17
Desmoglein-4 (DSG4)	Q86SJ6	2.3	3.5	4.3	128	103	134	15	3.81	7.63	15 624	31 250	3.3	5	13
Dipeptidyl aminopeptidase-like protein 6 (DPP6)	P42658	3.0	3.7	4.2	126	103	124	15	31	31	250 000	1 000 000	3.9	6	24
DNA topoisomerase 2-beta (TOP2B)	Q02880	1.3	1.7	2.5	200	101	227	15	15	31	31 250	31 250	3.0	6	16
Ectonucleoside triphosphate diphosphohydrolase 2 (ENTPD2)	Q9Y5L3	0.9	1.3	1.6	114	103	103	15	1.91	7.63	15 624	31 250	3.3	6	13
Ectonucleoside triphosphate diphosphohydrolase 6 (ENTPD6)	Q75354	2.4	2.6	3.0	121	104	117	15	244	244	62 499	499 999	2.4	4	22
EGF-like repeat and discoidin-like domain-containing protein 3 (EDIL3)	Q43854	1.5	2.1	3.2	113	100	100	1.9	61	61	62 499	1 000 000	3.0	5	19
Enteropeptidase (TMPRSS15)	P98073	0.0	0.9	2.4	131	101	121	15	15	15	31 250	31 250	3.3	3	20
Epidermal growth factor-like protein 7 (EGFL7)	Q9UHF1	4.2	4.9	6.0	133	108	135	15	61	61	124 999	250 000	3.3	5	19
Erythropoietin (EPO)	P01588	NA	0.9	2.8	131	104	126	15	244	244	62 499	124 999	2.4	6	19
Fatty acid-binding protein 9 (FABP9)	Q02758	1.1	1.8	2.4	118	102	117	15	15	15	31 250	124 999	3.3	5	22
Forkhead box protein O1 (FOXO1)	Q12778	1.0	1.5	2.3	74	103	69	15	244	488	499 999	499 999	3.0	5	22
Fructose-2,6-bisphosphatase TIGAR (TIGAR)	Q9NQ88	1.2	1.6	2.1	118	99	112	15	31	31	250 000	250 000	3.9	5	20
Hematopoietic prostaglandin D synthase (HPGDS)	Q60760	1.5	1.9	2.6	116	98	117	15	122	122	124 999	1 000 000	3.0	6	23
Inactive tyrosine-protein kinase 7 (PTK7)	Q13308	1.5	1.9	2.3	114	96	107	15	122	122	499 999	499 999	3.6	6	25
Integrin beta-1-binding protein 1 (ITGB1BP1)	Q14713	NA	NA	0.8	NA	NA	NA	15	244	244	250 000	499 999	3.0	7	21
Interferon-inducible double-stranded RNA-dependent protein kinase activator A (PRKRA)	Q75569	0.5	0.9	1.5	100	95	98	15	1953	3906	499 999	499 999	2.1	5	18
Kidney Injury Molecule (KIM1)	Q96D42	7.0	7.8	8.9	128	114	127	15	0.48	0.95	15 624	31 250	4.2	5	14
Leukotriene A-4 hydrolase (LTA4H)	P09960	NA	1.4	3.4	NA	NA	NA	15	7812	7812	499 999	499 999	1.8	4	30
Linker for activation of T-cells family member 2 (LAT2)	Q9GZY6	1.6	2.8	3.8	57	86	51	15	31	61	250 000	499 999	3.6	7	20
Lutropin subunit beta (LHB)	P01229	1.2	2.2	4.2	114	99	110	15	976	1953	1 000 000	1 000 000	2.7	5	24
Macrophage erythroblast attachor (MAEA)	Q7L5Y9	0.3	0.9	1.6	NA	NA	NA	3.8	61	61	62 499	124 999	3.0	6	18
Macrophage-capping protein (CAPG)	P40121	2.2	2.8	3.8	147	96	142	15	244	244	499 999	499 999	3.3	6	18
Melanoma-associated antigen D1 (MAGED1)	Q9Y5V3	NA	0.4	1.1	NA	NA	NA	15	15	15	15 624	31 250	3.0	5	13
Methionine aminopeptidase 1 (METAP1)	P53582	NA	NA	0.8	NA	NA	NA	15	244	244	250 000	499 999	3.0	4	21
Mevalonate kinase (MVK)	Q03426	1.8	2.7	4.6	143	110	137	15	15	31	62 499	250 000	3.3	5	19
Mitogen-activated protein kinase kinase kinase kinase 5 (MAP4K5)	Q9Y4K4	1.1	2.3	4.3	31	104	30	0.5	7.63	7.63	15 624	31 250	3.3	5	20

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		10th %tile	Median	90th %tile	ACD	Heparin	Serum	Hemolysate	LOD	LLOQ	ULOQ	Hook	Range	Intra	Inter
Mothers against decapentaplegic homolog 1 (SMAD1)	Q15797	NA	0.9	1.7	NA	105	NA	15	15	31	250 000	1 000 000	3.9	6	16
NAD-dependent protein deacylase sirtuin-5, mitochondrial (SIRT5)	Q9NXA8	NA	0.2	1.1	NA	NA	NA	15	488	488	250 000	499 999	2.7	6	16
NEDD8 ultimate buster 1 (NUB1)	Q9Y5A7	NA	0.9	1.8	95	99	NA	0.5	15	15	15 624	31 250	3.0	5	15
Neutrophil cytosol factor 2 (NCF2)	P19878	2.7	4.1	5.6	577	105	581	15	1.91	3.81	31 250	31 250	3.9	5	21
Nibrin (NBN)	Q60934	0.3	1.3	1.9	181	104	175	15	31	31	124 999	499 999	3.6	7	16
Nitric oxide synthase, endothelial (NOS3)	P29474	0.7	1.8	2.6	65	90	NA	15	7.63	15	62 499	124 999	3.6	5	16
Nucleobindin-2 (NUCB2)	P80303	3.1	3.6	4.3	173	107	170	15	31	31	62 499	124 999	3.3	5	24
Parvalbumin alpha (PVALB)	P20472	3.4	4.3	5.6	144	110	145	15	3.81	7.63	7812	31 250	3.0	4	18
Paxillin (PXN)	P49023	2.4	2.9	3.7	208	100	193	3.8	122	244	124 999	250 000	2.7	6	24
Peptidyl-prolyl cis-trans isomerase FKBP1B (FKBP1B)	P68106	NA	0.9	1.5	NA	NA	NA	15	122	244	62 499	124 999	2.4	5	19
Perilipin-1 (PLIN1)	Q60240	1.7	2.8	4.2	163	107	157	15	31	61	62 499	250 000	3.0	5	19
Phosphatidylinositol 3,4,5-trisphosphate 5-phosphatase 2 (INPPL1)	Q15357	1.3	2.1	3.6	NA	99	NA	3.8	122	122	62 499	499 999	2.7	5	18
Placenta growth factor (PGF)	P49763	8.0	8.5	9.0	133	111	130	15	0.48	0.95	7812	15 624	3.9	5	10
Platelet-derived growth factor C (PDGFC)	Q9NRA1	2.7	3.5	4.0	101	104	99	15	61	122	15 624	124 999	2.1	4	18
Pleiotrophin (PTN)	P21246	0.7	1.3	2.6	NA	NA	NA	15	244	244	31 250	31 250	2.1	7	15
Plexin domain-containing protein 1 (PLXDC1)	Q8IU55	0.4	1.4	1.8	108	106	NA	15	61	61	62 499	124 999	3.0	4	20
Polypeptide N-acetylgalactosaminyltransferase 10 (GALNT10)	Q86SR1	1.9	2.2	2.6	128	101	123	15	976	976	250 000	499 999	2.4	4	23
Probetaceinulin (BTC)	P35070	0.1	0.9	1.5	127	NA	119	15	0.95	7.63	7812	15 624	3.0	6	10
Programmed cell death protein 1 (PDCD1)	Q15116	1.1	1.3	1.6	NA	96	100	15	7.63	7.63	31 250	31 250	3.6	4	21
Prolow-density lipoprotein receptor-related protein 1 (LRP1)	Q07954	1.8	2.2	2.7	NA	98	NA	15	244	244	499 999	1 000 000	3.3	6	29
Proteasome subunit alpha type-1 (PSMA1)	P25786	NA	0.9	3.6	108	NA	NA	3.8	488	1953	250 000	499 999	2.1	4	18
Protein amnionless (AMN)	Q9BXJ7	2.3	3.4	4.3	128	105	122	15	3.81	15	31 250	124 999	3.3	4	26
Protein enabled homolog (ENAH)	Q8N8S7	1.7	2.3	3.1	127	109	117	15	61	61	62 499	62 499	3.0	5	17
Protein fosB (FOSB)	P53539	NA	0.8	2.4	NA	NA	NA	15	61	244	499 999	499 999	3.3	8	17
Protein LAP2 (ERBB2IP)	Q96RT1	0.7	1.9	3.3	NA	96	NA	0.5	61	61	31 250	499 999	2.7	4	22
Protein max (MAX)	P61244	NA	0.6	1.6	NA	NA	NA	15	122	122	62 499	124 999	2.7	5	18
Protein phosphatase 1B (PPM1B)	Q75688	NA	0.4	0.9	NA	NA	NA	7.5	244	244	499 999	499 999	3.3	5	17
Pyruvate dehydrogenase [acetyl-transferring]-phosphatase 1, mitochondrial (PDP1)	Q9POJ1	NA	NA	NA	NA	NA	NA	15	7812	7812	499 999	1 000 000	1.8	9	33
Ras association domain-containing protein 2 (RASSF2)	P50749	0.6	1.0	2.0	163	97	157	15	7.63	15	62 499	124 999	3.6	4	17
Ras GTPase-activating protein 1 (RASGAP1)	P20936	NA	NA	1.4	NA	NA	NA	15	NA	NA	NA	NA	NA	8	22
Receptor-type tyrosine-protein phosphatase eta (PTPRJ)	Q12913	NA	NA	1.2	127	92	127	15	488	976	499 999	1 000 000	2.7	6	23
Renin receptor (ATPGAP2)	Q75787	0.7	1.0	1.4	139	NA	124	15	NA	NA	NA	NA	NA	10	23
REST corepressor 1 (RCOR1)	Q9UKL0	0.8	1.0	1.5	127	NA	112	15	3.81	7.63	62 499	124 999	3.9	6	11
Retinoic acid receptor responder protein 1 (RARRES1)	P49788	1.7	2.1	2.5	92	98	NA	15	61	61	499 999	1 000 000	3.9	5	20
Ribonucleoside-diphosphate reductase subunit M2 B (RRM2B)	Q7LG56	0.3	0.9	1.3	142	102	136	15	7.63	15	499 999	1 000 000	4.5	12	14
Serpin A9 (SERPINA9)	Q86WD7	2.0	3.1	4.3	135	116	146	7.5	1.91	1.91	31 250	31 250	4.2	5	17
Serum paraoxonase/arylesterase 2 (PON2)	Q15165	2.5	2.9	3.6	82	102	74	15	122	122	124 999	499 999	3.0	9	23
Syntaxin-8 (STX8)	Q9UNK0	1.2	1.9	2.5	124	97	118	0.5	31	31	31 250	124 999	3.0	4	18
Syntaxin-binding protein 3 (STXB3)	Q00186	NA	NA	1.2	NA	NA	NA	0.9	61	61	124 999	250 000	3.3	5	20
Troponin I, cardiac muscle (TNNI3)	P19429	NA	1.5	5.4	NA	105	NA	15	61	122	499 999	499 999	3.6	10	19
Tyrosine-protein kinase Fes/Fps (FES)	P07332	NA	NA	0.7	NA	NA	NA	15	61	488	250 000	1 000 000	2.7	6	20
Tyrosine-protein kinase Fgr (FGR)	P09769	1.3	2.1	2.9	148	95	133	0.9	976	1953	499 999	499 999	2.4	5	22
Tyrosine-protein kinase Yes (YES1)	P07947	2.8	4.1	5.5	62	101	59	0.2	3.81	7.63	15 624	31 250	3.3	5	18
Vascular endothelial growth factor C (VEGFC)	P49767	1.3	1.7	2.7	310	NA	271	15	122	244	62 499	124 999	2.4	5	18
Vasohibin-1 (WASH1)	Q7L8A9	NA	NA	NA	NA	NA	NA	15	122	244	124 999	250 000	2.7	6	18
Wiskott-Aldrich syndrome protein (WAS)	P42768	NA	NA	1.4	NA	NA	NA	15	976	976	124 999	1 000 000	2.1	6	27

\*U/μl

## 2.3 PRECISION

### REPEATABILITY

Intra-assay variation (within-run) was calculated as the mean %CV for 6 individual samples run in triplicates within each of 6 separate runs during the validation studies. Inter-assay variation (between runs) was calculated between experiments with the same operator. The reported inter-assay %CV is the average of three operators' %CV. Variation calculations were performed on linearized values for 91 analytes for which response levels could be measured in serum and normal plasma, see Table 1.

Across all 91 assays, the mean intra-assay and inter-assay variations were observed to be 6.0% and 20.2%, respectively. The distribution of both intra-assay and inter-assay variations are shown in Figure 4.

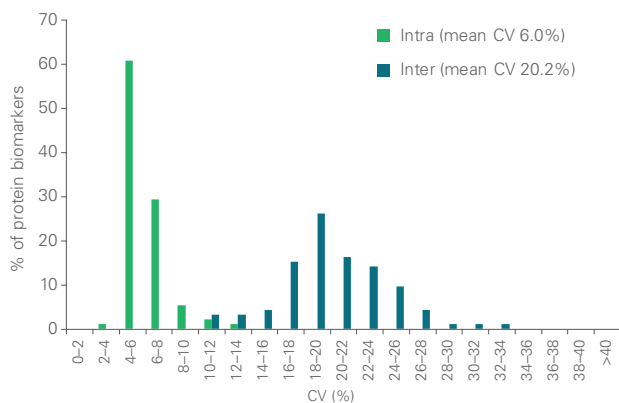


Fig 4. Distribution of intra-assay and inter-assay variations of Olink Organ Damage.

### REPRODUCIBILITY

Inter-site variations (between-site) were investigated during the validation of previous panels in beta-site studies to estimate the expected variations in values between different laboratories, with different operators and using different equipment. The beta-site studies have previously shown reproducibility and repeatability in line with Olink Proteomics results. For information on performed beta-site studies, download our Data Validation documents or contact support@olink.com.

## 2.4 ANALYTICAL SPECIFICITY

### ASSAY SPECIFICITY

The antibodies used in Olink Organ Damage were all specific for their respective targets. In principle, the specificity is tested by creating a test sample, consisting of a pool of antigens, which is then incubated with all 92 antibody probe pairs from the panel. Only if there is a correct match will a reporter sequence be created and serve as a template for subsequent real-time qPCR. Ten sub-pools of antigen are evaluated to cover the 92 assays in Olink, see Figure 5.

### ENDOGENOUS INTERFERENCE

Endogenous interference from heterophilic antibodies, e.g. human anti-mouse antibody (HAMA), and rheumatoid factor are known to cause problems in some immunoassays. Evaluation of the potential

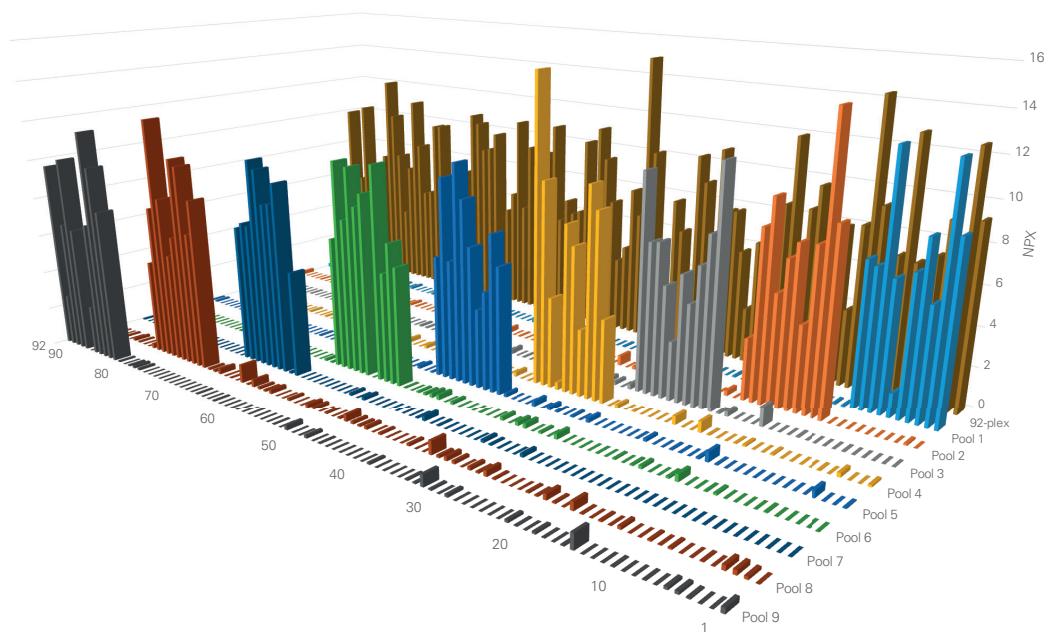
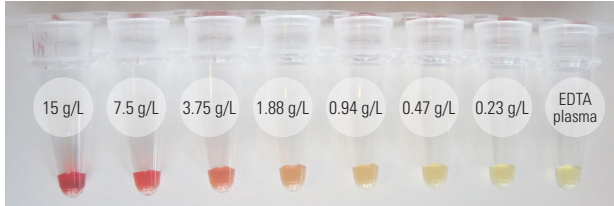


Fig 5. Assay readout specificity of the Olink platform. For each assay, specificity is confirmed by testing antigen sub-pools against the complete 92-plex pool as to each sub-mix.

impact of this specific interference was investigated during the validation of previous panels. No interference due to HAMA or RF could be detected for any of the samples in previously tested panels, indicating sufficient blocking of these agents (data not shown).

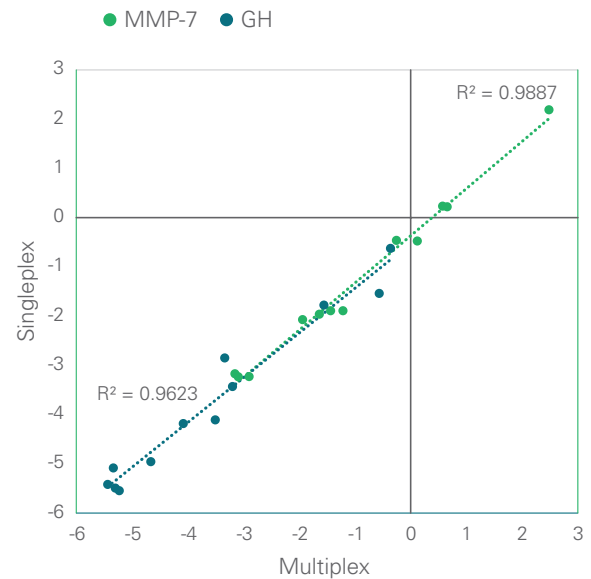


**Fig 6.** Endogenous interference. Levels tested for hemolysate were 0.23-15 g/L hemoglobin. The highest hemolysate concentration translates to about 10% hemolysis.

The potential impact of bilirubin, lipids and hemolysate, known interfering plasma and serum components, were evaluated at different added concentrations. An example of hemolysate levels tested is shown in Figure 6. These additions represent different patient health conditions and/or sample collection irregularities. Interferens by bilirubin and lipids has previously been evaluated, and disturbance has only been observed at extrem levels corresponding to 8 or 10 times normal <sup>3,4</sup> values and therefore not performed for Olink Organ Damage. In 16 out of 92 assays, altered signal was observed by the addition of hemolysate. The reason is most likely due to actual analyte leaking out of the disrupted blood cells. A concentration of 15 g/L of hemolysate represents 10% hemolysis of a sample. Table 1 reports the highest concentration of hemolysate that does not have an impact on assay performance.

## 2.5 SCALABILITY

Assay performance was further evaluated with regard to scalability, meaning the capability of the Olink technology to maintain the same quality of performance irrespective of multiplex level. Previously, we have shown that a step-wise increase of multiplex grade (8, 24, 48, 72 and 96) does not compromise assay performance (data not shown). To further strengthen that Olink provides consistent results, single assays for Growth Hormone (GH) and Matrix Metalloproteinase (MMP-7) were compared when run in a full 96-plex reaction. The results for each assay and their observed dCq-values were plotted against the entire 96-plex reaction. The square of the correlation coefficient ( $R^2$ ) value was generated by linear regression.



**Fig 7.** Scalability of the Olink technology platform. The experiment was performed using the Olink CVD II <sup>96x96</sup> panel. Human plasma samples were analyzed in singleplex for Growth Hormone (GH) and Matrix Metalloproteinase (MMP-7) with the equivalent assays performed in a full 96-plex reaction. The observed dCq (log2) values were plotted, and the correlation coefficient  $R^2$  value was generated by linear regression.



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## TECHNICAL SUPPORT

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1055, v2.0, 2018-03-16